1) Publication number:

0 184 187

EUROPEAN PATENT APPLICATION

- (1) Application number: 85115311.4
- (2) Date of filing: 03.12.85

(12)

(9) Int. Ct.*: C 12 N 15/00 C 07 K 13/00, C 12 N 5/00

- 30 Priority: 04.12.84 JP 254980'84
- (4) Date of publication of application: 11.06.86 Bulletin 86/24
- Designated Contracting States: CH DE FR GB LI
- Applicant: TEIJIN LIMITED
 11 Minamihonmachi 1-chome Higashi-ku
 Osaka-shi Osaka 541(JP)
- 72 Inventor: Kudo, Akira 3-1-1, Maedashi Higashi-ku Fukuoka-shi Fukuoka-ken(JP)
- (72) Inventor: Nishimura, Yushi 4-29, Chiyo-machi Hakata-ku Fukuoka-shi Fukuoka-ken(JP)
- 2 Inventor: Ichikawa, Yataro 2-11-7, Kotesashi-cho Tokorozawa-shi Saitama-ken(JP)
 - (2) Inventor: Watanabe, Takeshi 870-46, Yaemizo Nabeshima-cho Saga-shi Sakan-ken(JP)
- Representative: Kraus, Walter, Dr. et al, Patentanwälte Kraus, Welsert & Partner Thomas-Wimmer-Ring 15 D-8000 München 22(DE)

Mouse-human chimaeric immunoglobulin haavy chain, and chimaeric DNA encoding it.

(E) A mouse-human chimaeric immunoglobulin heavy chan comprising lat he anino aceli sequence of a mouse immunoglobulin heavy chain variable region and (b) the anino acid sequence of a human immunoglobulin heavy chair constant region and reacting specifically with human common acute lymphocytic levenile anigen and a chimaeric DNA fragment which encodes the amino acid sequtary changes of the control o

This invention relates to a mouse-human chimaeric immunoglobulin heavy chain and a chimaeric DNA sequence encoding it. More specifically, it relates to a mouse-human chimaeric immunoglobulin heavy chain comprising a combination of the variable region of a mouse immunoglobulin heavy chain and the constant region of a human immunoglobulin 10 heavy chain, and to a chimaeric DNA sequence encoding it.

Recent studies on immunoglobulin genes have provided much information on the heavy chains ("H-chain" for short) of immunoglobulin genes of mammals, particularly humans. It is well known that generally the H-chain is 15 composed of a variable region which has a site of binding to an antigen and a constant region which has physiological activities such as complement fixation or interaction with particular cells (see "Gendai Kagaku" (Modern Chemistry), November 1981, pages 62-68, published by Baifukan, Tokyo).

On the other hand, the DNA sequences of mouse immunoglobulin genes and their functions have been considerably elucidated. It is now known that a V-segment gene, a D-segment gene and a J-segment gene are aligned in this sequence in the variable region of mouse immunoglobulin genes and a DNA sequence having a promoter function exists upstream of the V-segment gene (5'-terminus). Furthermore. the existence of an enhancer, i.e. a DNA sequence having the function of greatly increasing transcriptional efficiency, between the J-segment gene and the constant region is pre-30 sumed. It is said that this enhancer exists downstream of

20

the J gene; when it is of the undifferentiated type, it cannot enhance transcription efficiency because the promoter is remote; but it becomes functionable upon recombination in V-J and V-D-J joinings and gains the function of enhancing 35 the efficiency of transcription from the promoter existing

upstream of the V gene.

Recently, Walter Schaffner et al. reported the DNA sequence of an enhancer in mouse immunoglobulin H-chain qenes (Cell, Vol. 33, 729-740, July 1983).

Susumu Tonegawa et al. likewise determined the existence of an enhancer in mouse immunoglobulin H-chain genes and its DNA sequence (see Cell, vol. 33, 717-728, July 1983).

David Baltimore et al. reported the existence of an enhancer in the light chains ("L-chain" for short) of 10 mouse immunoglobulin genes and its DNA sequence (see Cell, vol. 33, 741-748, July 1983).

The prior efforts have thus resulted in determining the existence of various gene segments and DNA segments having various functions, such as a promoter and an en-

15 hancer, in the variable and constant regions of mouse immunoglobulin H-chain and L-chain genes, and the DNA sequences of some of them.

On the other hand, there have been gradual elucidations of the structures and functions of constituent
genes of human immunoglobulin H-chain. But the existence of an enhancer was only presumed, and there was no knowledge of the entire DNA sequence of a human enhancer and DNA segments capable of expressing its function.

Very reently, however, T. H. Rabbitts et al. and 25 S. Tonegava et al. independently reported the DNA sequence of an enhancer in the H-chain of a human immunoglobulin gene (see Nature, vol. 306, 806-809, 22/2a, December 1983 and Nature vol. 307, 334-340, 26 January 1984).

Research on immunoglobulins has thus reached such
an advanced stage, and recently allowed utilization of
monoclonal antibodies which are expected to find applications in immunological diagnosis and therapy, for example in
the diagnosis, control and prevention of diseases. The use
of human immunoglobulins is most suitable in such immuno-

35 logical diagnosis and therapy utilizing such monoclonal antibodies, but human hybridoma techniques have not reached such a stage that human monoclonal antibodies having any required specificities can be easily obtained as in the case of mouse hybridomas. This imposes great restrictions on immunological diagnosis and therapy of human subjects. A method which might overcome this difficulty is to create artificially an antibody which is very similar to a human antibody and is not recognized as a foreign substance within a human body, and utilize this antibody for immunological diagnosis and therapy. As such an immunoglobulin, a chimaeric immunoglobulin composed of the variable regions of a certain mouse immunoglobulin and the constant regions of a human immunoglobulin was proposed very recently (European Patent Application EP 0 125 023 Al).

The present inventors have also worked extensively in this field in order to create a novel chimaeric immunoglobulin which can be applied to the immunological diagnosis and therapy of humans, and have now found a mouse-human chimaeric immunoglobulin heavy chain which specifically react with human common acute lymphocytic leukemia antigen (to be referred to as human "cALL" antigen) in combination with a proper immunoglobulin light chain.

According to this invention, there is provided a mouse-human chimaeric immunoglobulin heavy chain comprising (a) the amino acid sequence of a mouse immunoglobulin heavy chain variable region and (b) the amino acid sequence of a human immunoglobulin heavy chain constant region and reacting specifically with human common acute lymphocytic leukemia antigen in combination with a proper immunoglobulin light chain.

30 Human cALL antigen is an antigen commonly observed in the bloods of patients with various types of acute lymphocytic leukemia. A monoclonal antibody which specifically reacts with human cALL antigen but is not recognized as a foreign substance by humans would greatly contribute the 35 immunological diagnosis and therapy of patients with acute lymphocytic leukemia.

The immunoglobulin heavy chain of this invention has been developed to meet such a desire. It is characterized by having a chimnaeric structure composed of

- (a) the amino acid sequence of a mouse immuno-
- 5 globulin heavy chain variable region, and
 - (b) the amino acid sequence of a human immunoglobulin heavy chain constant region, and also immunologically by reacting specifically with human cALL antigen.

The variable region of the mouse immunoglobulin

10 heavy chain, as mentioned hereinabove, consists roughly of a

V-segment, a D-segment and a J-segment, and it is the Vsegment which is most involved in the recognition of antigens. The D-segment and J-segment exist in various types
depending upon their amino acid sequences, and for example,

5 the existence of J₁, J₂, J₃ and J₄ in the J-segment is known. The D- and/or J-segment of the mouse H-chain variable region in the present invention is not limited to any particular amino acid sequence but may vary so long as an antibody finaly constructed from the H-chain of this in-

20 vention has the property of specifically reacting with human cALL antigen. Preferably, however, an H-chain variable region derived from a mouse immunoglobulin H-chain reacting specifically with human cALL antigen is suitable as the mouse immunoglobulin H-chain variable region used in this

25 invention. Specific examples of this variable region include a variable region whose V-segment has the following amino acid sequence

Asp Val Gln Leu Val Glu Ser Gly Gly Cly Leu Val
Gln Pro Gly Gly Ser Arg Lys Leu Ser Cys Ala Ala
30 Ser Gly Phe Thr Phe Ser Ser Phe Gly Met His Trp
Val Arg Gln Ala Pro Glu Lys Gly Leu Glu Trp Val
Ala Tyr Ile Ser Gly Gly Ser Tyr Thr Ile Tyr Tyr
Ala Asp Thr Val Lys Gly Arg Phe Thr Ile Ser Arg
Asp Asn Pro Lys Asn Thr Leu Phe Leu Gln Met Thr
35 Ser Leu Arg Ser Glu Asp Thr Ala Met Tyr Tyr Cys
Ala Ser Ser Tyr Gly Asn Phe Trp Tyr Phe Asp Val

Trp Gly Ala Gly Thr Thr Val Thr Val Ser Ser and more specifically a variable region having the following total amino acid sequence

Asp Val Gln Leu Val Glu Ser Gly Gly Gly Leu Val

Gln Pro Gly Gly Ser Arg Lys Leu Ser Cys Ala Ala
Ser Gly Phe Thr Phe Ser Ser Phe Gly Met Bis Trp
Val Arg Gln Ala Pro Glu Lys Gly Leu Glu Trp Val
Ala Tyr Ile Ser Gly Gly Ser Tyr Thr Ile Tyr Tyr
Ala Asp Thr Val Lys Gly Arg Phe Thr Ile Ser Arg
Asp Asn Pro Lys Asn Thr Leu Phe Leu Gln Met Thr
Ser Leu Arg Ser Glu Asp Thr Ala Met Tyr Tyr Cys
Ala Ser Ser Tyr Gly Asn Phe Trp Tyr Phe Asp Val
Trp Gly Ala Gly Thr Thr Val Thr Val Ser Ser Ser
Tyr Gly Asn Phe Trp Tyr Phe Asp Val Trp Gly Ala
Gly Thr Thr Val Thr Val Ser Ser.

In the present and served.

In the present applications, the following abbreviations are used for various amino acids.

Gly: glycine Ala: alanine

Val: valine

20

25

30

35

Leu: leucine

Ile: isoleucine

Ser: serine

Asp: aspartic acid Lys: lysine

Arg: arginine

His: histidine

Phe: phenylalanine

Tyr: tyrosine

Thr: threonine Cvs: cysteine

Met: methionine

Glu: glutamic acid

Trp: tryptophan

Pro: proline

Asn: asparagine

Gln: glutamine

ferred.

On the other hand, polypeptides constituting the constant region of the chimaeric immunoglobulin H-chain of this invention may be those derived from the H-chain of any of IgA, IgD, IgE, IgG and IgM which are known as human immunoglobulins (Ig). Generally, those derived from IgG, above all IgG, are suitable. More specifically, a polypeptide having the following amino acid sequence is pre-

Ala Ser Thr Lys Gly Pro Ser Val Phe Pro Leu Ala 10 Pro Ser Ser Lys Ser Thr Ser Gly Gly Thr Ala Ala Leu Gly Gys Leu Val Lys Asp Tyr Phe Pro Glu Pro Val Thr Val Ser Trp Asn Ser Gly Ala Leu Thr Ser Gly Val His Thr Phe Pro Ala Val Leu Gln Ser Ser Gly Leu Tyr Ser Leu Ser Ser Val Val Thr Val Pro 15 Ser Ser Ser Leu Gly Thr Gln Thr Tyr Ile Gys Asn Val Asn His Lys Pro Ser Asn Thr Lys Val Asp Lys Lys Val Glu Pro Lys Ser Cys Asp Lys Thr His Thr Cys Pro Pro Cys Pro Ala Pro Glu Leu Leu Gly Gly Pro Ser Val Phe Leu Phe Pro Pro Lys Pro Lys Asp 20 Thr Leu MET Ile Ser Arg Thr Pro Glu Val Thr Cvs Val Val Val Asp Val Ser His Glu Asp Pro Glu Val Lys Phe Asn Trp Tyr Val Asp Gly Val Glu Val His Asn Ala Lys Thr Lys Pro Arg Glu Glu Gln Tyr Asn Ser Thr Tyr Arg Val Val Ser Val Leu Thr Val Leu 25 His Gln Asp Trp Leu Asn Gly Lys Glu Tyr Lys Cys Lys Val Ser Asn Lys Ala Leu Pro Ala Pro Ile Glu Lys Thr Ile Ser Lys Ala Lys Gly Gln Pro Arg Glu Pro Gln Val Tyr Thr Leu Pro Pro Ser Arq Glu Glu MET Thr Lys Asn Gln Val Ser Leu Thr Cys Leu Val 30 Lys Gly Phe Tyr Pro Ser Asp Ile Ala Val Glu Trp Glu Ser Asn Gly Gln Pro Glu Asn Asn Tyr Lys Thr Thr Pro Pro Val Leu Asp Ser Asp Gly Ser Phe Phe Leu Tyr Ser Lys Leu Thr Val Asp Lys Ser Arg Trp Gln Gln Gly Asn Val Phe Ser Cys Ser Val MET His 35 Glu Ala Leu His Asn His Tyr Thr Gln Lys Ser Leu Ser Leu Ser Pro Gly Lys.

One typical mouse-human chimaeric immunoglobulin H-chain provided by this invention has the amino acid sequence shown in Fig. 1 of the accompanying drawings.

The chimaeric immunoglobulin H-chain of this

invention can be produced by joining a DNA fragment encoding
the amino acid sequence of the mouse immunoglobulin variable
region to a DNA fragment encoding the amino acid sequence of
the human immunoglobulin constant region to construct a
chimaeric DNA fragment capable of expression, inserting the
chimaeric fragment into a plasmid vector, and introducing
the recombinant plasmid vector into host animal cells to
cause the chimaeric DNA fragment to express in the host
cells.

The DNA fragment encoding the amino acid sequence

15 of the mouse immunoglobulin variable region can be obtained,
for example, by preparing hybridomas by cell fusion between
spleen cells taken from a mouse immunized with membrane
pieces of human cALL and mouse myeloma cells, and screening
the chromosomal DNA of the hybridomas (see, for example, T.

20 Maniatis et al., Molecular Cloning, Cold Spring Harbor Lab., 1982). Specific procedures therefor are described in detail in Examples 9 to 11 given hereinbelow.

On the other hand, the DNA fragment encoding the amino acid sequence of the human immunoglobulin constant
25 region can be obtained by screening the chromosomal DNA of cells producing human immunoglobulins by a conventional method. Specific procedures therefor are described in detail in Examples 1 to 3 given hereinbelow.

Joining of the two DNA fragments described above 30 through a DNA fragment (to be referred to as the "hhsegment") at least containing a human enhancer gives a chimaeric DNA sequence encoding the mouse-human chimaeric immunoglobulin H-chain. The Eh-segment should at least contain a human enhancer. It may additionally contain a 35 mouse enhancer or other internal additionally contain a

35 mouse enhancer or other introns derived from a human or mouse.

The chimaeric DNA fragment is inserted into a suitable plasmid vector such as pSV2gpt, pSV2neo and pKSV-10 to prepare a recombinant plasmid vector.

The recombinant plasmid vector so constructed is 5 introduced into E. coli for example. Thereafter, the recombinant plasmid vector is introduced into host animal cells such as mouse myeloma J558L or NS-1 by the protoplast fusion method (see "Men-eki Jikken Sosa Ho" (Operational Methods in Immunological Experiments) XIII, p. 4533, edited 10 by the Japanese Society of Immunology, 1984). By culturing the resulting cells, the desired mouse-human chimaeric immunoglobulin H-chain can be accumulated in, and recovered from, the culture.

The chimaeric H-chain provided by this invention

15 may be combined with a mouse-human chimaeric immunoglobulin
light chain prepared, for example, by the same technique as
described above to form a complete mouse-human chimaeric
immunoglobulin. Such an immunoglobulin is expected to be
used in the immunological treatment of cALL by administering

20 it to human cALL patients either alone or in combination
with an anticancer agent.

The following Examples more specifically illustrate the construction and expression of a chimaeric DNA sequence encoding the chimaeric H-chain of this invention.

25 Example 1

Isolation of a human chromosomal DNA:Human cultivated cells ARB77 (3 x 10 cells) were
crushed by a glass rod and treated with Protease K (a product of Sigma Co.) in the presence of 2% sodium dodecylsulfate (SDS). Then, phenol saturated with 10mM Tris HCl
(pH 8.0)-lmM EDTA aqueous solution was added. The mixture
was centrifuged to separate it into an aqueous layer and a
phenol layer. The aqueous layer was dialyzed against 20mM
Tris HCl (pH 7.5)-100mM NaCl-5mM EDTA aqueous solution. The
dialyzate was treated with Ribonuclease A (a product of
Sigma Co.) and after phenol extraction, dialyzed against

10mM Tris HCl (pH 8.0)-lmM EDTA aqueous solution to obtain about 1.2 mg of a human chromosomal DNA [see N. Blin and D. W. Stafford, Nucleic Acids Res., 3, 2303 (1976)]. Example 2

Preparation of a human gene library:-The human chromosomal DNA (150 micrograms) obtained in Example 1 was partially digested with restriction endonuclease EcoRI (produced by Takara Shuzo Co., Ltd.) in acccordance with the method shown in Example 4, and then 10 subjected to sucrose density gradient centrifugation [sucrose 10-40% (wt/vol), 28,000 rpm x 15 hours, 20°C] to obtain 4.3 micrograms of a DNA fragment having a size of 15 Kb to 23 Kb. The DNA fragment (0.8 microgram) was ligated by using T4-DNA ligase (a product of Bethesda Research 15 Laboratory) with Charon 4A vector [see F. R. Blattner, B. G. Williams, A. E. Blechl, K. D. Thompson, H. E. Faber, L. A. Furlong, D. J. Grunward, D. O. Kiefrr, D. O. Moore, J. W. Schumm, F. L. Sheldon and Smithies, Science, 196, 161 (1977)] to obtain a hybrid DNA in which the human DNA was 20 inserted between the right arm and left arm of Charon 4A vector. The ligating reaction was carried out at 11°C for 12 hours in 66mm Tris-HCl (pH 7.6)-6.6mm MgCl2-10mm dithiothreitol-1 mm ATP aqueous solution. The resulting hybrid DNA was packaged in vitro [A. Becker and M. Gold, 25 Proc. Natl. Acad. Sci. U. S., 72, 581 (1975)] to form a human gene library (1.8 x 10 PFU/µg-DNA, containing more than 99% of human chromosomal DNA).

Example 3 Screening of human immunoglobulin H-chain gene:-E. coli LE 392 strain (ATCC 33572) was infected with the gene library (an assembly of Charon 4A phages containing human chromosomal DNA) obtained in Example 2 to form a plaque. Clones containing human immunoglobulin gene were selected in accordance with the plaque hybridization 35 method of Benton and Davis [see W. D. Benton and R. W. Davis, Science, <u>196</u>, 180 (1977)] using a ³²p-labelled human

30

immunoglobulin H-chain J gene. DNA was prepared from the Charon 4A phages containing human immunoglobulin gene by the method of Thomas and Davis [M. Thomas and R. W. Davis, J. Mol. Biol., 91, 315 (1974)].

Example 4

tion.

Preparation of a restriction endonuclease cleavage map of human immunoglobulin gene:-One microgram of the Charon 4A DNA containing a human immunoglobulin gene obtained in Example 3 was dis-10 solved in 20 microliters of a buffer for digestion with restriction endonucleases [50mm Tris-HCl (pH 7.4)-100mm NaCl-10mM MgSO, aqueous solution for EcoRI, EcoRV, MluI or SphI; 10mm Tris-HC1 (pH 7.5)-60mm NaCl-7mm MgCl, aqueous solution for AatII, AvaI, BamHI, BstEII, HincII, HindIII, 15 PstI or PvuII; 10mM Tris-HC1 (pH 7.4)-10mM MgSO4-lmM dithiothreitol aqueous solution for BglII; and 10mM Tris-ECl (pH 8.0)-20mm KC1-7mm MgCl2-7mm 2-mercaptoethanol solution for HpaI], and digested with two units of a restriction endonuclease (AatII was a product of Toyobo Ltd.; AvaI was a 20 product of Bethesda Research Laboratories, Inc.; BstEII and $\underline{\operatorname{Sph}}I$ were products of New England Biolabs. Inc.; and the other endonucleases were products of Takara Shuzo Co., Ltd.) at 37°C for more than 1 hour. Cleavage with <u>Bst</u>EII was carried out at 60°C for more than 1 hour. When the diges-25 tion was carried out by using two restriction endonucleases, there was used a procedure comprising first treating the DNA with one restriction endonuclease acting at a low salt concentration, then increasing the salt concentration to a

After the digestion, 4 microliters of an aqueous solution containing 0.25% of bromophenol blue and 50% of glycerol, and the mixture was subjected to 0.8%-2.5% agarose gel electrophoresis. The agarose used was type II made by Sigma Co. designed for electrophoresis. A 40mM Tris-CH₂COOH

predetermined concentration, and treating DNA with the other restriction endonuclease acting at a higher salt concentra(pH 8.0)-lmM EDTA aqueous solution was used as an electro-phoresis running buffer. With a vertical gel having a thickness of 2 mm, the electrophoresis was carried out at a voltage of 6 to 9 V/cm for 1.5 to 3 hours. A product of digestion of λ phage DNA with HindIII (made by Boeling

- of digestion of λ phage DNA with <u>Hin</u>dIII (made by Boeling Manheim) was used as a molecular size marker for DNA fragments. After the electrophoresis, the DNA in the agarose gel was dyed with a 2 micrograms/ml aqueous solution of ethidium bromide, and by irradiating long wavelength ultra-
- 10 violet light onto the gel, the digestion pattern was observed. By analyzing the patterns of digestion with the various restriction endonucleases either alone or in a combination of two, the relative positions of the restriction endonuclease cleavage sites were determined.

15 Example 5

Recloning of a human immunoglobulin gene (a hybrid DNA of a 8.2 Kb <u>Hin</u>dIII fragment containing a human C_{γ1} gene and <u>E. coli</u> plasmid pBR322):Three micrograms of Charon 4A DNA containing a

- 20 human immunoglobulin gene was digested with BindIII substantially in accordance with the method of Example 4, and subjected to agarose gel electrophoresis (gel concentration 0.8%). A band corresponding to DNA having a size of 8.2 kb and containing a C₁ gene was cut out and the agarose gel
- 25 slice was dissolved in three times (vol/wt) its amount of an 8M aqueous NaClO₄ solution. A DNA fragment having a size of 8.2 kb was recovered from the agarose gel by the glass filter method of Chen and Thomas [C. W. Chen and C. A. Thomas Jr., Anal. Biochem. 101, 339 (1980)].
- Separately, 1 microgram of <u>E. coli</u> plasmid pBR322 was digested with <u>BindIII</u> in accordance with Example 4, and reacted with 0.5 unit of alkaline phosphatase (<u>E. coli</u> C75) (a product of Takara Shuzo Co., Ltd.) at 45°C for 1 hour. After the reaction, the reaction mixture was extracted with
- 35 water saturated phenol three times in order to deactivate and remove the alkaline phosphatase in the reaction mixture.

The alkaline phosphatase treated product of the above digested pBR322 was mixed with an aqueous solution of the 8.2 kb <u>Hind</u>III fragment recovered from the agarose gel. The mixture was precipitated with ethanol, and the pre-

- 5 cipitate was dissolved in 50 microliters of a buffer for ligation (see Example 2). Two units of T4-DNA ligase was added, and the reaction was carried out at 11°C for 12 hours to give a hybrid DNA.
- E. coli C600 strain (ATCC 33525) was transformed 10 with the hybrid DNA by an improved version of the usual CaCl₂ method [M. V. Norgard, K. Keen and J. J. Monaham, Gene. 3, 297 (1978)]. Specifically, E. coli C600 strain cultivated for 18 hours was inoculated in 5 ml of L-broth (1% tryptone, 0.5% yeast extract, 0.5% NaCl, pH 7.2) and
- 15 grown to an extent corresponding to an optical density at 600 nm of 0.3. The cells were washed twice in a cold magnesium buffer [0.1M NaCl-5mM MgCl₂-5mM Tris-HCl (pH 7.6, 0°C)], re-suspended in 2 ml of a cold calcium buffer [100mM CaCl₂-250mM KCl-5mM MgCl₂-5 mM Tris-HCl (pH 7.6, 0°C)], and
- 20 left to stand at 0°C for 25 minutes. The cells were then concentrated to one-tenth of their volume in a calcium buffer, and mixed with an aqueous solution of the hybrid DNA in a ratio of 2:1 (vol:vol). The mixture was maintained at 0°C for 60 minutes, and after adding 1 ml of LBG-broth (1%
- 25 tryptone, 0.5% yeast extract, 1% NaCl, 0.08% glucose, pH 7.2), cultivated at 37°C for 1 hour. The culture fluid was inoculated in a selective medium (an L-broth plate containing 30 micrograms/ml of ampicillin) at a rate of 100 microliters/plate. The plate was cultivated overnight at 37°C
- 30 to grow the transformants. A DNA was prepared from the resulting colony by a known method, and by agarose gel electrophoresis, the existence of the desired hybrid DNA was determined.

Restriction endonuclease cleavage map of subclones

35

restriction endonuclease cleavage maps shown in Figs. 2 and 3 of the accompanying drawings were obtained.

Fig. 2 shows fragments obtained by cleaving a human chromosomal DNA containing an immunoglobulin gene with 5 restriction endonuclease EcoRI. The 5'-terminal position is indicated as EcoRI-(1), and the 3'-terminal position, as EcoRI-(2). Fig. 2 also shows sites at which the DNA fragment can be cleaved with restriction endonuclease XbaI and XhoI. This fragment was cleaved with some restriction 10 eondonucleases and divided into several small fragments. Some fragments were inserted into the E. coli plasmid vector pBR322 to obtain four subclones shown in Fig. 3.

Subclone pTJlB:

Obtained by inserting a small DNA fragment (about 15 8.2 kb) interposed between HindIII-(2) and HindIII-(3) into the HindIII site of pBR322. The restriction endonuclease cleavage site map of this subclone is shown in Fig. 3, II-A. The presence of 3 to 4 Pst I sites were recognized between PstI-(3) and HindIII-(3) in Fig. 3, II-A. 20

Subclone pTJ1BR:

25

Obtained by insertion into pBR322 in the same way as in the preparation of pTJlB above, but the orientation is reverse to that in pTJlB. The restriction endonuclease cleavage map of this subclone is shown in Fig. 3, II-B.

Subclone pTJ5: Obtained by inserting a small DNA fragment existing between PstI-(2) and PstI-(3) of the above subclone pTJlB into the PstI site of pBR322. The restriction endonuclease cleavage map of this subclone is shown in Fig. 3, 30 II-A-(1).

Subclone pTJ4:

Obtained by deleting from the above subclone pTJlB a portion ranging from a Pst I site closest to Hind III-(3) existing between Hind III-(3) and Pst I-(3) to Pst I-(2). 35 The restriction endonuclease cleavage map of this subclone is shown in Fig. 3, II-A-(2).

- 14 -

Subclone pTJ3:

Obtained by inserting a small DNA fragment (about 3.4 kb) interposed beween EcoRI-(1) and BindIII-(1) into a site between EcoRI and HindIII. A detailed restriction sendonuclease cleavage map of this subclone is shown in Fig. 3, I.

Example 6

Determination of the DNA sequence of the human immunoglobulin gene enhancer region:-

The DNA sequences of the enhancer region and the V-D region of the H-chain of a human immunoglobulin gene were determined by the Maxam-Gilbert method [A. Maxam and W. Gilbert, Methods Enzymol., 65, 499 (1980)].

About 50 micrograms of the subclone pTJ3 DNA

- 15 prepared in Example 5 was digested with <u>Bst</u>EII in accordance with the method of Example 4. The resulting DNA fragment was dephosphorylated with alkaline phosphatase, and labelled with [7-32] ATP using 5 units of polynucleotidekinase (produced by P-L Bjochemicals Co.). The polynucleo-
- 20 tidekinase reaction was carried out in a 50mM Tris-HCl (pH 9.5)-10mM MgCl₂-5mM dithiothreitol aqueous solution, and [x-3²P] ATP (made by Amersham Co.) was used in an amount corresponding to 100 micro Ci. The ³²P-labelled DNA fragment was digested with BpaI. The desired DNA fragment
- 25 was separated by polyacrylamide gel electrophoresis (gel concentration 5%) and extracted from the gel. The resulting ³²P-labelled <u>BstEII/HpaI</u> fragment was subjected to a partial hydrolysis reaction specific for each of the bases, and subjected to electrophoresis on polyacrylamide gel con-
- 30 taining 7M urea (gel concentration 8% to 23%). The separated fragment was autoradiographed at -80°C for 2 to 7 days, and the cleavage pattern was analyzed to obtain data for the determination of the DNA sequence of the enhancer region.
- 35 Thus, the DNA sequences including the enhancer DNA sequence of this invention shown in Fig. 4 were determined.

Example 7

Determination of the DNA base sequence of a human immunoglobulin gene constant region:-The base sequence of the constant region of a

5 human immunoglobulin gene was determined by the Maxam-Gilbert method [A. Maxam and W. Gilbert, Methods Enzymol., 65, 499 (1980)].

pTJ5 was digested with Pst1, and labelled with [a.32p] ddATP using a 3'-terminal labelling kit (made by 10 Amersham Co.). The 32p-labelled DNA fragment was digested with Sma1, and the desired DNA fragment was separated and recovered by polyacrylamide gel elecrophoresis (gel concentration 5%). The resulting 32p-labelled Pst1/Sma1 DNA fragment was subjected to a partial hydrolysis reaction 15 specific for each of the bases, and separated by electrophoresis on polyacrylamide gel containing 7M urea (gel concentration 8% to 23%). Then, the DNA fragment was autoradiographed at -80°C for 2 to 7 days. The cleavage pattern was then analyzed to obtain data for the determination of the DNA sequence of the constant region as shown in Fig. 5.

Example 8

Construction of a human immunoglobulin gene expression plasmid:-

25
An EcoRI fragment of the human H-chain gene (HIGI) obtained in Example 3, with a size of about 21 Kbp, was inserted into the EcoRI site of vector pSV-2gpt developed by P. Berg [R. C. Mulligan and P. Berg, Proc. Natl. Acad. Sci., U.S.A., 78, 2072 (1981)] to obtain a plasmid pSV2-HIGI The restriction endonuclease cleavage map of this plasmid is shown in Fig. 6.

Example 9

Isolation of a mouse chromosomal DNA:
Mouse hybridoma NL-1 cells (2 x 10⁷) obtained from

Dr. R. Ueda of Laboratory of Chemotherapy, Aichi Cancer

Center Research Institute, Nagoya 464, Japan were treated

with protease K in the presence of 1% SDS. Water-saturated phenol was then added to extract DNA. The aqueous layer was separated by centrifugation and dialyzed against 10mM Tris-BCl buffer (pB 7.4) containing 0.1mM NaCl and 0.1mM EDTA (TNE buffer). The aqueous solution was treated with ribonuclease A, and again extracted with water-saturated phenol. The aqueous layer was dialyzed against the TNE buffer to obtain 300 micrograms of mouse chromosomal DNA. Example 10

Preparation of a mouse gene library:-

10

35

The mouse chromosomal DNA (150 micrograms) obtained in Example 9 was completely digested with restriction endo-nuclease EcoRI and subjected to agarose gel electrophoresis. A DNA fragment (5 micrograms) corresponding to 7 kb-9 kb was 15 recovered from the agarose gel by electroelution.

The resulting DNA fragment (0.4 microgram) and Charon 4A vector <u>EcoRI</u> arms (Amersham Co.) were ligated by means of T4 DNA ligase, and packaged in-vitro using a kit of Amersham Co. As a result, a mouse hybridoma NL-1 gene 20 libraray (8 x 10⁶ PFU/microgram) was obtained. Example 11

Screening of mouse immunogloblin H-chain gene:
E. Coli strain LE 392 was infected with the Charon

4A phage obtained in Example 10 containing DNA derived from

mouse hybridoma NL-1 to form a plaque. Clones containing

mouse antibody H-chain genes were screened by the plaque
hybridization method using a 32P-labelled mouse immunogloblin H-chain J gene.

This procedure led to the isolation of a gene
including all V regions (a 5' flanking region, VDJ regions
and an enhancer region) of mouse hybridoma NL-1 having a
size of 7.9 kb.
Example 12

Preparation of a restriction endonuclease Cleavage map:-

A 7.9 kb EcoRI fragment of the mouse hybridoma

NL-1 H-chain DNA obtained in Example 11 was recloned into vector pBR322 and used to transform <u>E. Coli</u> strain DL-1 (ATCC 33849). The <u>E. Coli</u> strain was cultivated in large quantities to obtain about 1 mg of a plasmid (pBR-NL-1-H)

- 5 having the 7.9 kb mouse hybridoma NL-1 H chain DNA fragment inserted into the EcoRI site of pBR322. The pBR NL-1-H was cleaved with restriction endonucleases, <u>ECoRI</u>, <u>BamHI</u>, <u>HindIII</u>, <u>EcoRV</u>, and <u>PvuII</u> (Takara Shuzo Co., Ltd.) and <u>SphI</u> (Bethesda Research Laboratory), and a restriction endo-
- 10 nuclease cleavage map was prepared as shown in Fig. 7. As a typical example, cleavage with <u>Eco</u>RI is shown below. <u>Example 13</u>

Determination of the DNA sequence of mouse immunoglobulin VDJ region:-

The pBR-NL-1-H was cleaved with restriction endonucleases <u>Sph</u>I and <u>PvuII</u> to obtain two DNA fragments given by the inserted gene between <u>Sph</u>I and <u>PvuII</u> sites. The DNA fragments were cloned into M13 phase vectors mpl8 and mpl9 (P. L. Biochemicals Co.) cleaved with <u>Sph</u>I and <u>Hin</u>CII. DNA 20 sequence determination was performed by the dideoxy chain

termination method using an M13 sequencing kit (Takara Shuzo Co., Ltd.). The sequence of the SphI/PvuII fragment on the 5' side was determined both in the downstream direction from the SphI site and in the upstream direction from the PvuII

25 site. The sequence of the <u>SphI/PvuII</u> fragment on the 31 side was determined in the downstream direction from the <u>PvuII</u> site. The DNA sequence of the variable region is shown in Fig. 8.

Example 14

Construction of chimaeric immunoglobulin gene expression plasmid pMH-1:-

The plasmid pBR-NL-1-H obtained by inserting an anti-human cALL immunoglobulin H-chain gene fragment (7.9 kb) into the EcoRI site of plasmid vector pBR322 was cleaved with restriction endonucleases EcoRV and BamHI. An EcoRV and BamHI fragment (about 2.7 kb) containing the entire mouse

immunoglobulin H-chain variable region was separated by agarose gel electrophoresis and purified by the method of Example 3.

In the meantime, the plasmid vector pSV2-gpt 5 mentioned hereinabove was cleaved with restriction endonucleases BamHI and HpaI, and a linear vector fragment having a size of about 5.3 kb and containing the Ecogpt gene was separated and purified (this vector gene is abbreviated as "fragment-1").

10

The EcoRV-BamHI fragment (about 2.7 kb) of pBR-NL-1-H obtained above was ligated with the BamI-HpaI fragment (about 5.3 kb) of pSV2-gpt mentioned above by using T4-DNA ligase. Subsequent cleavage with restriction endonuclease BamHI gave a linear DNA fragment (about 8.0 kb) containing 15 about 2.7 kb of the variable region of NL-1-H. The DNA fragment was separated and purified (this DNA fragment gene

is abbreviated as "fragment-2"). Purthermore, plasmid pSV-2-HIG1 containing a human immunoglobulin gene (about 21 kb) was cleaved with restric-

- 20 tion endonuclease MluI to give a DNA fragment (about 17 kb) containing a human enhancer and $C_{\sim 1}$, which was separated and A BamHI linker (a product of Takara Shuzo Co., Ltd.) was ligated with the resulting gene fragment (about 17 kb) in accordance with the method of T. Maniatis et al.
- 25 (T. Maniatis et al., "Molecular Cloning, Cold Spring Harbor Lab., 1982) to give a BamHI site to both ends (the resulting product is abbreviated as "fragment-4").

The fragment-4 was ligated with the fragment-2 by using T4-DNA ligase. E. coli strain MC1000 was transformed 30 with the ligation product to obtain a chimaeric immunoglobulin gene in which the variable region of the mouse origin was joined to the constant region of the human origin through the human enhancer (Eh) (this gene is abbreviated as "pMH-1"). The flow chart for the preparation of the plasmid pMH-1 is shown in Fig. 9.

Example 15

Construction of chimaeric immunoglobulin gene expression plasmid pMH-2:-

- Plasmid pSV-2-HIG1 containing a human immuno-5 globulin H-chain gene (about 21 kb) was cleaved with restriction endonuclease BamHI by a conventional method to give a fragment (about 14 kb) containing a human enhancer (Eh) and C v1. The resulting fragment was ligated with the fragmen: -2 by using T4-DNA ligase. E. coli MC1000 was
- 10 transformed with the ligation product to give a chimaeric immunoglobulin gene in which the variable region was of the mouse origin and the constant region and the enhancer were of the human origin (this plasmid is abbreviated as "pMH-2"). The flow chart for the preparation of the plasmid
- 15 pMH-1 is shown in Fig. 9.

Example 16

Construction of chimaeric immunoglobulin gene expression plasmid pMH-3:-

- The plasmid pBR-NL-1-H was cleaved with restric-20 tion endonucleases EcoRV and EcoRI in the same way as in
- Example 3 to give a fragment (about 4.7 kb) containing the entire mouse immunoglobulin H-chain variable regions and the mouse enhancer (Em). Then, the vector pSV-2-gpt was cleaved with restriction endonucleases EcoRI and HpaI to give a
- 25 fragment (about 4.7 kb) containing Ecogpt gene, which was separated and purified. The resulting fragment was ligated with the above mouse V gene fragment (about 4.7 kb) by using T4-DNA ligase, and the ligation product was then cleaved with restriction endonuclease EcoRI to give a linear DNA 30 fragment (about 9.2 kb) containing the variable region
- (about 4.7 kb) of NL-1-H, which was separated and purified (this DNA fragment is abbreviated as "fragment-3").
- Plasmid pSV2-HIG1 containing a human immunoglobulin H-chain gene (about 21 kb) was cleaved with 35 restriction endonuclease <u>Mlu</u>I to give a fragment (about 17 kb) containing a human enhancer (Eh) and C_{v1} , which was

separated and purified. An <u>EcoRI</u> linker (a product of Takara Shuzo Co., Ltd.) was ligated with the resulting fragment (about 17 kb) in accordance with the method of Maniatis et al. described in "Moclecular Cloning" to give 5 an <u>EcoRI</u> site to both ends (the resulting fragment is abbreviated as "fragment-5").

The fragment-5 was ligated with the fragment 3 by using T4-DNA ligase to give a chimaeric immunoglobulin gene having the human and mouse enhancers, a variable region of the mouse origin and a constant region of the human origin (this plasmid is abbreviated as "DMH-3").

The flow chart for the preparation of the plasmid pMH-3 is shown in Fig. 9. Example 17

15

Construction of a chimaeric immunoglobulin gene expression plasmid pMH-4:-

The plasmid pBR-NL-1-H was cleaved with restriction endonucleases <u>Eco</u>RV and <u>Bam</u>HI to give a fragment (about 2.7 kb) containing the entire V-gene segment of a mouse,

0 which was separated and purified. <u>EcoRI</u> linker was ligated with the <u>EcoRV</u> terminus of this fragment to give an <u>EcoRI</u> site and form an EcoRI-BamHI fragment (about 2.7 kb).

In the meantime, the vector pSV2-gpt was cleaved with EcoRI and BamHI to give a linear vector fragment (4.6 to containing an Ecogpt gene, which was separated and purified. The resulting vector fragment was ligated with the above EcoRI-BamHI fragment (about 2.7 kb) by using T4-DNA ligase. The ligation product was then cleaved with restriction endonuclease BamHI to give a DNA fragment (about 30 7.3 kb) containing the variable region gene of NL-1-H (this DNA fragment is abbreviated as "fragment-6").

The fragment-6 was ligated with the fragment-4 obtained in Example 14 containing the human enhancer (Eh) and C₇₁ by using T4-DNA ligase. E. coli MC1000 was transformed with the ligation product to give a chimaeric immunoglobulin gene having a variable region of the mouse

origin and a constant region of the human origin joined through the human enhancer (Eh) (this plasmid is abbreviated as "pMH-4").

The flow chart for the preparation of the plasmid 5 $\,$ pMH-4 is shown in Fig. 10.

Example 18

Construction of chimaeric immunoglobulin gene expression plasmid pMH-5:-

The plasmid pBR-NL-1-H was cleaved with restriction endonucleases BamHI and EcoRI to obtain a fragment
(about 2 kb) containing a mouse enhancer (Em), which was
separated and purified. The fragment (about 2 kb) was
ligated with the fragment-5 obtained in Example 16 by using
T4-DNA ligase. The ligation product was cleaved with restriction endonuclease BamHI to give a BamHI fragment (about
13 kb) containing the mouse and human enhancers and human
C. ... The BamHI fragment was historia with a single containing the mouse and human containing the mouse a

C_{v1}. The <u>Bam</u>HI fragment was ligated with the fragment-6 obtained in Example 17 by using T4-DNA ligase. <u>E. coli</u> MC1000 was transformed with the ligation product to give a chimaeric immunoglobulin gene containing a variable region of the mouse origin and a constant region of the human

of the mouse origin and a constant region of the human origin joined through the mouse and human enhancers (this plasmid is abbreviated as "pMH-5").

The flow chart for the construction of the plasmid $25\,$ pMH-5 is shown in Fig. 10.

Example 19

Expression of the chimaeric immunoglobulin genes:-

Each of the chimáeric immunoglobulin gene expression plasmids pMH-1 to pMH-5 obtained in Examples 14 to 18
was introduced into <u>E. coli</u> MCl000 in accordance with the
method described in Example 5. The transformants were
cultured in a medium containing ampicillin, and the plasmid
was amplified by adding chloramphenicol. <u>E. Coli</u> proto-

35 plasts were obtained by treatment with lysozyme (Sigma Co.). The resulting protoplasts were fused with 2 \times 10 mouse

myeloma cells J558L or NS-l in 50% PEG 4000 for 2 to 7 minutes. The fusion product was diluted with MEM medium (Gibco, Grand Island, New York), and PEG 4000 was removed by centrifugation. The resulting cells were grown for 48 to 72 5 hours in RPMI 1640 complete medium (Gibco). The cell surfaces were then stained by fluorescence using PITC-labelled goat anti-human IgG (Cappel Laboratories, Inc.), and the

goat anti-human IgG (Cappel Laboratories, Inc.), and the expression of the chimaeric immonogloblin gene was checked.

Strong expression was observed in the cells into which the expression plasmids pMH-1, pMH-2 and pMH-4 were introduced.

expression plasmids pMH-1, pMH-2 and pMH-4 were introduced in On the other hand, relatively weak expession was observed in the cells into which the expression plasmids pMH-3 and pMH-5 were introduced.

RNA was extracted from the individual cells, and
15 20 micrograms of the resulting RNA was subjected to northern
hybridization (T. Maniatis et al., Molecular Cloning, Cold
Spring Harbor Lab., 1982) using a <u>BamHI-PvuII</u> fragment of
plasmid pBR-NL-1-H containing mouse V, D, and J genes and a
<u>PstI(-(2)-PstI-(3)</u> fragment of plasmid pSV2-HIGl containing
20 part of human C₇₁ gene as probes. Strong hybridization was
observed with RNAs of the cells in which the expression
plasmids pMH-1, pMH-2 and pMH-4 were introduced, whereas
hybridization was relatively weak with the RNAs of the cells
in which the expression plasmids pMH-3 and pMH-5 were in25 troduced.

In the present example, the degree of expression showed the same tendency in mouse myeloma J558L and NS-1.

The expression plasmid pMH-4 was introduced into mouse myeloma cells J558L by protoplast fusion and trans30 ferred to a selective medium containing 250 \(\gamma/m\) xanthine,
15 \(\gamma/m\) hypoxanthine and 6 \(\gamma/m\) mycophenolic acid. The
medium was exchanged for two successive days. Two weeks
later, transformants having resistance to mycophenolic acid
were obtained. mRNA was extracted from the transformants,
35 and subjected to northern hybridization using a V-gene of a
mouse VDJ and a human C, gene as a probe. At a size of

- 23 **-**

1.7 kb, a band was observed which corresponded to mRNA of a complete secretor ${\tt H-chain}$.

The transformants were cultured in the presence of \$35\$S-methionine (Amersham). The supernatant obtained was immunoprecipitated with anti-human IgG. The precipitate was subjected to polyacrylamide gel electrophoresis and fluorographed. It was determined that the chimaeric immunoglobulin H-chain protein was present in the form of \$H_2L_2\$ in the supernatant of the culture.

Southern hybridization was carried out in an attempt to detect the chimaeric immunoglobulin gene. This led to the determination that the chimaeric antibody gene existed in an average copy number of about 0.5 per cell.

What is claimed is:

- A mouse-human chimaeric immunoglobulin heavy chain comprising (a) the amino acid sequence of a mouse immunoglobulin heavy chain variable region and (b) the amino acid sequence of a human immunoglobulin heavy chain constant region and reacting specifically with human common acute lymphocytic leukemia antigen.
- The chimaeric immunoglobulin heavy chain of claim l wherein the amino acid sequence of the variable region is derived from a mouse immunoglobulin heavy chain which reacts specificaly with human common acute lymphocytic leukemia antigen.
- 3. The chimaeric immunoglobulin heavy chain of claim 1 wherein the amino acid sequence of the constant region is derived from the heavy chain of a human immunoglobulin G.
- 4. The chimaeric immunoglobulin heavy chain of claim 3 wherein the human immunoglobulin G is human immunoglobulin G_1 .
- 5. The chimaeric immunoglobulin heavy chain of claim 1 wherein the variable region contains a V-segment having the following amino acid sequence

Asp Val Gln Leu Val Glu Ser Gly Gly Gly Leu Val Gln Pro Gly Gly Ser Arg Lys Leu Ser Cys Ala Ala Ser Gly Phe Thr Phe Ser Ser Phe Gly Met His Trp Val Arg Gln Ala Pro Glu Lys Gly Leu Glu Trp Val Ala Tyr Ile Ser Gly Gly Ser Tyr Thr Ile Tyr Tyr Ala Asp Thr Val Lys Gly Arg Phe Thr Ile Ser Arg Asp Asn Pro Lys Asn Thr Leu Phe Leu Gln Met Thr Ser Leu Arg Ser Glu Asp Thr Ala Met Tyr Tyr Cys Ala Ser Ser Tyr Gly Asn Phe Trp Tyr Phe Asp Val Trp Gly Ala Gly Thr Thr Val Thr Val Ser Ser Ser

wherein the various abbreviations stand for the following amino acids:

Gly: glycine
Ala: alanine
Val: valine

Leu: leucine

Ile: isoleucine

Ser: serine

Asp: aspartic acid

Lys: lysine

Arg: arginine

Bis: histidine

Phe: phenylalanine

Tyr: tyrosine

Thr: threonine

Cys: cysteine

Met: methionine

Glu: glutamic acid

Trp: tryptophan Pro: proline

Asn: asparagine

Gln: glutamine.

The chimaeric immunoglobulin heavy cyhain of claim 5 wherein the variable region has the following amino acid sequence

Asp Val Gln Leu Val Glu Ser Gly Gly Gly Leu Val Gln Pro Gly Gly Ser Arg Lys Leu Ser Cys Ala Ala Ser Gly Phe Thr Phe Ser Ser Phe Gly Met His Trp Val Arg Gln Ala Pro Glu Lys Gly Leu Glu Trp Val Ala Tyr Ile Ser Gly Gly Ser Tyr Thr Ile Tyr Tyr Ala Asp Thr Val Lys Gly Arg Phe Thr Ile Ser Arg Asp Asn Pro Lys Asn Thr Leu Phe Leu Gln Met Thr Ser Leu Arg Ser Glu Asp Thr Ala Met Tyr Tyr Cys Ala Ser Ser Tyr Gly Asn Phe Trp Tyr Phe Asp Val Trp Gly Ala Gly Thr Thr Val Thr Val Ser Ser Ser Tyr Gly Asn Phe Trp Tyr Phe Asp Val Trp Gly Ala Gly Thr Thr Val Thr Val Ser Ser

wherein the abbreviations for the amino acids are as shown in claim 5.

The chimaeric immunoglobulin heavy chain of claim 1 wherein the constant region has the following amino acid sequence

Ala Ser Thr Lys Gly Pro Ser Val Phe Pro Leu Ala Pro Ser Ser Lys Ser Thr Ser Gly Gly Thr Ala Ala Leu Gly Gys Leu Val Lys Asp Tyr Phe Pro Glu Pro Val Thr Val Ser Trp Asn Ser Gly Ala Leu Thr Ser Gly Val His Thr Phe Pro Ala Val Leu Gln Ser Ser Gly Leu Tyr Ser Leu Ser Ser Val Val Thr Val Pro Ser Ser Ser Leu Gly Thr Gln Thr Tyr Ile Gys Asn Val Asn His Lys Pro Ser Asn Thr Lys Val Asp Lys Lys Val Glu Pro Lys Ser Cys Asp Lys Thr His Thr Cys Pro Pro Cys Pro Ala Pro Glu Leu Leu Gly Gly Pro Ser Val Phe Leu Phe Pro Pro Lys Pro Lys Asp Thr Leu MET Ile Ser Arg Thr Pro Glu Val Thr Cvs Val Val Val Asp Val Ser His Glu Asp Pro Glu Val Lys Phe Asn Trp Tyr Val Asp Gly Val Glu Val His Asn Ala Lys Thr Lys Pro Arg Glu Glu Gln Tyr Asn Ser Thr Tyr Arg Val Val Ser Val Leu Thr Val Leu His Gln Asp Trp Leu Asn Gly Lys Glu Tyr Lys Cys Lys Val Ser Asn Lys Ala Leu Pro Ala Pro Ile Glu Lys Thr Ile Ser Lys Ala Lys Gly Gln Pro Arg Glu Pro Gln Val Tyr Thr Leu Pro Pro Ser Arg Glu Glu MET Thr Lys Asn Gln Val Ser Leu Thr Cys Leu Val Lys Gly Phe Tyr Pro Ser Asp Ile Ala Val Glu Trp Glu Ser Asn Gly Gln Pro Glu Asn Asn Tyr Lys Thr Thr Pro Pro Val Leu Asp Ser Asp Gly Ser Phe Phe Leu Tyr Ser Lys Leu Thr Val Asp Lys Ser Arg Trp Gln Gln Gly Asn Val Phe Ser Cys Ser Val MET His

wherein the abbreviations for the amino acids are as shown in claim 5.

8. The chimaeric immunoglobulin heavy chain of claim 1 which has the following amino acid sequence

Asp Val Gln Leu Val Glu Ser Gly Gly Gly Leu Val Gln Pro Gly Gly Ser Arg Lys Leu Ser Cys Ala Ala Ser Gly Phe Thr Phe Ser Ser Phe Gly Met His Trp Val Arg Gln Ala Pro Glu Lys Gly Leu Glu Trp Val Ala Tyr Ile Ser Gly Gly Ser Tyr Thr Ile Tyr Tyr Ala Asp Thr Val Lys Gly Arg Phe Thr Ile Ser Arg

Asp Asn Pro Lys Asn Thr Leu Phe Leu Gln Met Thr Ser Leu Arg Ser Glu Asp Thr Ala Met Tyr Tyr Cys Ala Ser Ser Tyr Gly Asn Phe Trp Tyr Phe Asp Val Trp Gly Ala Gly Thr Thr Val Thr Val Ser Ser Ser Tyr Gly Asn Phe Trp Tyr Phe Asp Val Trp Gly Ala Gly Thr Thr Val Thr Val Ser Ser Ala Ser Thr Lys Gly Pro Ser Val Phe Pro Leu Ala Pro Ser Ser Lys Ser Thr Ser Gly Gly Thr Ala Ala Leu Gly Gys Leu Val Lys Asp Tyr Phe Pro Glu Pro Val Thr Val Ser Trp Asn Ser Gly Ala Leu Thr Ser Gly Val His Thr Phe Pro Ala Val Leu Gln Ser Ser Gly Leu Tyr Ser Leu Ser Ser Val Val Thr Val Pro Ser Ser Ser Leu Gly Thr Gln Thr Tyr Ile Gys Asn Val Asn His Lys Pro Ser Asn Thr Lys Val Asp Lys Lys Val Glu Pro Lys Ser Cys Asp Lys Thr His Thr Cys Pro Pro Cys Pro Ala Pro Glu Leu Leu Gly Gly Pro Ser Val Phe Leu Phe Pro Pro Lys Pro Lys Asp Thr Leu MET Ile Ser Arg Thr Pro Glu Val Thr Cys Val Val Val Asp Val Ser His Glu Asp Pro Glu Val Lys Phe Asn Trp Tyr Val Asp Gly Val Glu Val His Asn Ala Lys Thr Lys Pro Arg Glu Glu Gln Tyr Asn Ser Thr Tyr Arg Val Val Ser Val Leu Thr Val Leu His Gln Asp Trp Leu Asn Gly Lys Glu Tyr Lys Cys Lys Val Ser Asn Lys Ala Leu Pro Ala Pro Ile Glu Lys Thr Ile Ser Lys Ala Lys Gly Gln Pro Arg Glu Pro Gln Val Tyr Thr Leu Pro Pro Ser Arg Glu Glu MET Thr Lys Asn Gln Val Ser Leu Thr Cys Leu Val Lys Gly Phe Tyr Pro Ser Asp Ile Ala Val Glu Trp Glu Ser Asn Gly Gln Pro Glu Asn Asn Tyr Lys Thr Thr Pro Pro Val Leu Asp Ser Asp Gly Ser Phe Phe Leu Tyr Ser Lys Leu Thr Val Asp Lys Ser Arg Trp Gln Gln Gly Asn Val Phe Ser Cys Ser Val MET His Glu Ala Leu His Asn His Tyr Thr Gln Lys Ser Leu Ser Leu Ser Pro Gly Lys

wherein the abbreviations for the amino acids are as shown in claim 5.

- A chimaeric DNA fragment which encodes the amino acid sequence of the mouse-human chimaeric immunoglobulin heavy chain of claim 1.
- 10. The chimaeric DNA fragment of claim 9 which contains a variable region V-segment DNA sequence represented by the following

GAT GTG CAG CTG GTG GAG TCT GGG GGA GGC TTA GTG
CAG CCT GGA GGG TCC CGG AAA CTC TCC TGT GCA GCC
TCT GGA TTC ACT TTC AGT AGC TTT GGA ATG CAC TGG
GTT CGT CAG GCT CCA GAG AAG GGG CTG GAC TGG
GCA TAT ATT AGT GGT GGC AGT TAT ACC ATC TAC TAT
GCA GAC ACA GTG AAG GGC CGA TTC ACC ATC TCC AGA
GAC AAT CCC AAG AAC ACC CTG TTC CTA CAA ATG ACC
AGT CTA AGG TCT GAG GAC ACG GCC ATG TAT TAC TGT
GCA AGT TCC TAT GGT AAC TTC TGG TAC TTC CAT
GGG GC GCA GGG ACC ACG GTC ACC GTC TCC TCA
Wherein A represents deoxyadenosine-5'-phosphate,
C represents deoxyquinosine-5'-phosphate, and T represents deoxythymidine-5'-phosphate,
and T repre-

and a DNA sequence complementary thereto.

11. The DNA sequence of the chimaeric DNA fragment of claim 10 which contains a variable region DNA sequence represented by the following

Wherein A, C, G and T are as defined in claim 10, and a DNA sequence complementary thereto.

- 12. The chimaeric DNA fragment of claim 9 which contains a human immunoglobulin heavy chain constant region DNA fragment comprising
- (1) a $C_{\rm H}1$ segment which contains DNA sequence represented by the following

(2) an h segment which contains a DNA sequence represented by the following

GAG CCC AAA TCT TGT GAC AAA ACT CAC ACA TGC CCA CCG TGC CCA

(3) a ${\rm C_{\rm H}}{\rm 2}$ segment which contains a DNA sequence represented by the following

GCA CCT GAA CTC CTG GGG GGA CCG TCA GTC TTC CTC
TTC CCC CCA AAA CCC AAG GAC ACC CTC ATG ATC TCC
CGG ACC CCT GAG GTC ACA TGC GTG GTG GTG GAC GTG
GTG GAC GCC GAG GTC ACA TGC GTC AAG TCC AAC AAC
CTG GAA GAC CCT GAG GTC AAA TG CC AAG ACA AAG
CCG CGG GAG GAG CAG TAC AAC AGC ACG TAC CGG GTG
GTC AGC GTC CTC ACC GTC CTG CAC CAG GAC TGG CTG
AAT GGC AAG GAG TAC AAG TGC AAG GTC TCC AAC AAA
GCC CTC CCA GCC CCC ATC GAG AAA ACC ATC TCC AAA

and (4) a $C_{\rm H}^{-3}$ segment which contains a DNA sequence represented by the following

GGG CAG CCC CGA GAA CCA CAG GTG TAC ACC CTG CCC CCA TCC CGG GAG GAG ATG ACC AAG AAC CAG GTC AGC CTG ACC TGC CTG GTC AAA GGC TTC TAT CCC AGC GAC ATC GCC GTG GAG TGG GAC AAC AAT GGG CAG CCG GAG AAC AAC TAC AAG ACC ACG CCT CCC GTG CTG GAC TCC GAC GAC GAC AAC AAC ACC TTC CTC TAT AGC AAG CTC ACC GTG GAC AAG AGC AGG TGG CAG CAG GGG AAC GTC TTC TCA TGC TCC GTG ATG CAT GAC GCT CTG CAC AAC CAC TAC ACG CAG AAG AAC ACC CT TAC ACG CAG AAG AAC ACC CAT TAC ACG CAG AAG AAC ACC CTC TCC CTG TCC CCG GGT AAA Wherein A, C, G and T are as defined in claim 10,

wherein A, C, G and T are as defined in claim 10, and a DNA sequence complementary to said constant region DNA sequences.

- 13. The chimaeric DNA fragment of claim 9 comprising a DNA fragment encoding the amino acid sequence of the variable region and a DNA fragment encoding the amino acid sequence of the constant region joined to each other through a DNA sequence containing at least a human enhancer.
- 14. The chimaeric DNA fragment of claim 13 wherein the human enhancer contains a DNA sequence represented by the following

TTG GCG AGC TGG AAG CAG ATG ATG AAT TAG AGT CAA GAT GCC TGC ATG GGG GTC TCC GGC ACC CAC AGC AGG TGG CAG GAA GCA AAA AAA CAC AAT TGG TAA ATT TAT CAC TTG TGA AGA GCT GGT TTT GCC AGG CCC AGA TCT GAA AGT GCT CTA CTG GAC AAA ACA CTT GGA CAA TTT GCG TTT TTT ACC AGG GCT GAC CAA ATT TGG TTT TTT AAC TAT CTG CAA ATT CAT TTC CAA ATG GCT TTT TAAC TAT CTG CAA TTT CAT TTC CAA TTT TAG CTT AAC TAT CTG CAA TTT GC

wherein A, C, G and T are as defined in claim 10, and a DNA sequence complementary thereto.

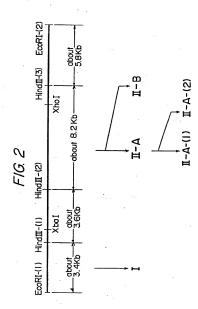
- 15. Recombinant pSV2gpt plasmid harboring the chimaeric DNA fragment of claim 9.
- 16. Mouse myeloma J558LK or NS-1 cells into which the chimaeric DNA fragment of claim 9 has been introduced by using the recombinant plasmid of claim 15.

1|11 FIG. 1-(1)

Asp Val Gln Leu Val Glu Ser Gly Gly Gly Leu Val Gln Pro Gly Gly Ser Arg Lys Leu Ser Cys Ala Ala Ser Gly Phe Thr Phe Ser Ser Phe Gly Met His Trp Val Arg Gln Ala Pro Glu Lys Gly Leu Glu Trp **V**al Als Tyr Ile Ser Gly Gly Ser Tyr Thr Ile Tyr Tyr Ala Asp Thr Val Lys Gly Arg Phe Thr Ile Ser Arg Asp Asn Pro Lys Asn Thr Leu Phe Leu Gln Met Thr Ser Leu Arg Ser Glu Asp Thr Ala Met Tyr Tyr Cys Ala Ser Ser Tyr Gly Asn Phe Trp Tyr Phe Asp Val Trp Gly Ala Gly Thr Thr Val Thr Val Ser Ser Ser Tyr Gly Asn Phe Trp Tyr Phe Asp Val Trp Gly Ala Gly Thr Thr Val Thr Val Ser Ser Ala Ser Thr Lys Gly Pro Ser Val Phe Pro Leu Ala Pro Ser Ser Lys Ser Thr Ser Gly Gly Thr Ala Ala Leu Gly Cys Leu Val Lys Asp Tyr Phe Pro Glu Pro Val Thr Val Ser Trp Asn Ser Gly Ala Len Thr Ser Gly Val His Thr Phe Pro Ala Val Leu Gln Ser Ser Gly Len Tyr Ser Leu Ser Ser Val Val Thr Val Pro Ser Ser Ser Leu Gly Thr Gln Thr Tyr Ile Cys Asn Val Asn His Lys Pro Ser Asn Thr Lys Val Asp Lys

FIG. 1- (2)

Lys Val Glu Pro Lys Ser Cys Asp Lys Thr His Thr Cys Pro Pro Cys Pro Ala Pro Glu Leu Leu Gly Gly Pro Ser Val Phe Leu Phe Pro Pro Lys Pro Lys Asp Thr Leu MET Ile Ser Arg Thr Pro Glu Val Thr Cys Val Val Val Asp Val Ser His Glu Asp Pro Glu Val Lys Phe Asn Trp Tyr Val Asp Gly Val Glu Val His Asn Ala Lys Thr Lys Pro Arg Glu Glu Gln Tyr Asn Ser Thr Tyr Arg Val Val Ser Val Leu Thr Val Leu His Gln Asp Trp Leu Asn Gly Lys Glu Tyr Lys Cys Lys Val Ser Asn Lys Ala Leu Pro Ala Pro Ile Glu Lys Thr Ile Ser Lys Ala Lys Gly Gln Pro Arg Glu Pro Gln Val Tyr Thr Leu Pro Pro Ser Arg Glu Glu MET Thr Lys Asn Gln Val Ser Leu Thr Cys Leu Val Lys Gly Phe Tyr Pro Ser Asp Ile Ala Val Glu Trp Glu Ser Asn Gly Gln Pro Glu Asn Asn Tyr Lys Thr Thr Pro Pro Val Leu Asp Ser Asp Gly Ser Phe Phe Leu Tyr Ser Lys Leu Thr Val Asp Lys Ser Arg Trp Cln Cln Cly Asn Val Phe Ser Cys Ser Val MET His Glu Ala Leu His Asn His Tyr Thr Gln Lys Ser Leu Ser Leu Ser Pro Gly Lys



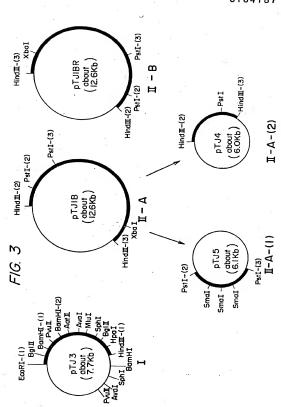


FIG. 4

FIG. 5-(1)

C_H1-segment;

GCCTCCACCAAGGGCCCATCGGTCTTCCCCCTGGCA
CCCTCCTCCAAGAGCACCTCTGGGGGCACAGCGGCC
CTGGGCTGCCTGGTCAGGGACTACTTCCCCGAACCG
GTGACGGTCTCGTGGAACTCAGGCGCCCTGACCAGC
GGCGTGCACACCTTCCCGGCTGTCCTACAGTCCTCA
GGACTCTACTCCCTCAGCAGCGTGGTGACCGTGCCC
TCCAGCAGCTTGGGCACCCAGACCTACATCTGCAAC
GTGAATCACAAGCCCAGCAACACCAAGGTGGACAAG
AAAGTT

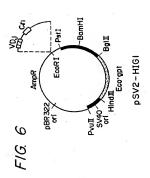
h-segment;
GAGCCCAAATCTTGTGACAAAACTCACACATGCCCA
CCGTGCCCA

C_H2-segment;

GCACCTGAACTCCTGGGGGGACCGTCAGTCTTCCTC
TTCCCCCCAAAACCCAAGGACACCCTCATGATCTCC
CGGACCCCTGAGGTCACATGCGTGGTGGTGGACGTG
AGCCACGAAGACCCTCAGGTCAAGTTCAACTGGTAC
GTCGACGCCTGGAGGTGCATAATGCCAAGACAAAG
CCGCGGGAGGACCAGTACAACAGCACGTACCGGGTG
GTCAGCGTCCTCACCGTCCTGCACCAGGACTGGCTG
AATGGCAAGGAGTACAAGTGCAAGGTCTCCAACAAA
GCCCTCCCAGCCCCCATCGAGAAAACCATCTCCAAA
GCCAAA

C_H3-segment;

GGGCAGCCCGAGAACCACAGGTGTACACCCTGCCC
CCATCCCGGGAGGAGCATGACCAAGAACCAGGTCAGC
CTGACCTGCCTGGTCAAAAGGCTTCTATCCCAGCGAC
ATCGCCGTGGAGTGGCAGAGCAATGGGCAGCCGGAG
AACAACTACAAGACCACGCCTCCCGTGCTGGACTCC
GACGGCTCCTTCTTCCTCTATAGCAAGCTCACCGTG
GACAAGAGCAGGTGGCAGCGGGAACGTCTTCTCA
TGCTCCGTGATGCATGAGGCTCTGCACAACCACTAC
ACGCAGAAGAGGCCTCTCCCCTGTCCCCGGGTAAA



EcoRI 7,9 (Kbp) EcoR_V . 5 EcoR_V HindII BamHI SphI PvuII SphI SphI 3.0 3.3 3.7 5.0 PvuI 0.3 EcoRI

FIG. 8

GATGTGCAGCTGGTGGAGTCTGGGGGAGGCTTAGTGCAGCCTV-segment-
GGAGGCTCCCGGAAACTCTCCTGTGCAGCCTCTGGATTCACT
TTCAGTAGCTTTGGAATGCACTGGGTTCGTCAGGCTCCAGAC
AAGGGGCTGGAGTGGCAGTTAT
ACCATCTACTATGCAGACACAGTGAAGGGCCGATTCACCATC
TCCAGAGA CAATCCCAAGAACACCCTGTTCCTACAAATGACC
AGTCTAAGGTCTGAGGACACGGCCATGTATTACTGTGCAAGT
TCCTATGGTACTTCTGGTACTTCGATGTCTGGGGCGCAGGG
ACCACGGTCACCGTCTCCTCATCCTATGGTAACTTCTGGTAC
TTCGATGTCTGCGGCGCAGGGACCACGGTCACCGTCTCCTCA
J ₁ -segment

Fig. 9

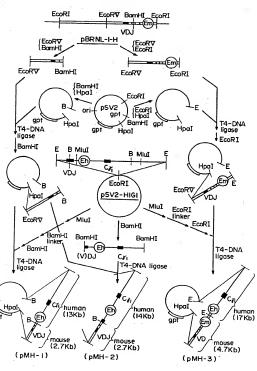


FIG. 10

